

## TWO CHEMICALLY DISTINCT GROUPS OF *CALOPHYLLUM* SPECIES FROM SRI LANKA

B. M. RATNAYAKE BANDARA, H. RANJITH W. DHARMARATNE, SUBRAMANIAM SOTHEESWARAN\* and SINNATHAMBY BALASUBRAMANIAM†

Department of Chemistry, University of Peradeniya, Peradeniya, Sri Lanka; †Department of Botany, University of Peradeniya, Peradeniya, Sri Lanka

(Revised received 11 June 1985)

**Key Word Index**—*Calophyllum* species; Guttiferae; (+)-*cis*-dihydroinophyllolide; inophyllum A; (–)-*trans*-dihydroinophyllolide; soulattrolide; calozeylanic acid.

**Abstract**—The leaf extracts of *Calophyllum moonii* and *C. walkeri* were investigated. The former contained neoflavonoids whereas the latter had calozeylanic acid. Distribution of natural products in the leaf extracts of seven *Calophyllum* species has been reviewed and the existence of two chemically distinct groups in *Calophyllum* species has been recognized.

### INTRODUCTION

Nine endemic *Calophyllum* species are recorded in Sri Lanka. They are: *C. bracteatum* Thw., *C. calaba* L., *C. cordato-oblongum* Thw., *C. cuneifolium* Thw., *C. lankaensis* Kosterm (= *C. zeylanicum* Kosterm.), *C. moonii* (= *C. soulattri* Burm f.), *C. thwaitesii* Planch and Triana, *C. trapezifolium* Thw. and *C. walkeri* Wight. We have reported [1–3] the chemical investigation of the bark and leaves of a few *Calophyllum* species. In this paper we report on the leaf extractives of two species, namely, *C. moonii* and *C. walkeri*. These, and our previous results, indicate the existence of two chemically distinct groups of *Calophyllum* species.

### RESULTS AND DISCUSSION

The hot petrol extracts of the leaves of *C. moonii* when chromatographed over silica gel and eluted with petrol–EtOAc gave (i) a sitosterol ester which on hydrolysis with potassium hydroxide gave sitosterol and (ii) D:A-friedo-oleanan-3-one (friedelin). Two neoflavonoids were eluted next and have been characterized as (+)-*cis*-dihydroinophyllolide (1) [4] and (–)-*trans*-dihydroinophyllolide (2) [5].

It appeared from the data given in the Experimental that 1 is a neoflavonoid and is probably (+)-*cis*-dihydroinophyllolide (inophyllum A). The following decoupling studies to confirm the stereochemistry at C-11 and C-10 were carried out on this neoflavonoid. Irradiation at  $\delta_H$  1.16 caused the multiplet at  $\delta_H$  2.27 to collapse to a double-doublet ( $J = 3.46$  and 5.19 Hz) and irradiation at  $\delta_H$  1.43 caused the multiplet at  $\delta_H$  4.43 to collapse to a doublet ( $J = 3.46$  Hz). The complete chemical shift data are given in Fig. 1. These results confirm that the neoflavonoid is identical to inophyllum A (1) charac-

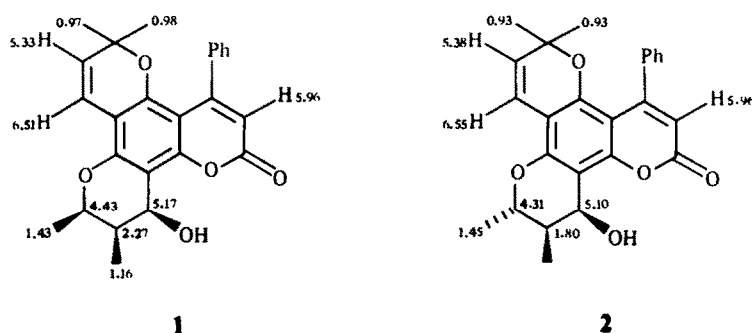
terized by Kawazu *et al.* [4, 6]. The data for 1 suggest that H-10–H-11 is *gauche* ( $J = 3.46$  Hz) and that H-11–H-12 is probably *gauche* ( $J = 5.19$  Hz). Thus the C-10 methyl and the hydroxyl are pseudo-equatorial and the C-11 methyl is pseudo-axial.

The second neoflavonoid isolated had *M*, (mass spectrum), mass spectral fragmentation pattern, IR and UV spectra all identical to 1. However, the  $[\alpha]_D$  value was negative and the  $^1H$  NMR spectrum was slightly different to that of 1. Irradiation at  $\delta_H$  1.18 resulted in the multiplet at  $\delta_H$  1.80 collapsing to a double doublet ( $J = 3.46$  and 10.30 Hz). Irradiation at  $\delta_H$  1.45 resulted in the multiplet at  $\delta_H$  4.31 collapsing to a doublet ( $J = 10.30$  Hz). The neoflavonoid is thus identified as soulattrolide (2) and it exists in a conformation in which both methyl groups at C-10 and C-11 are pseudo-equatorial and the hydroxyl group at C-12 is pseudo-axial. This leads to a *gauche* arrangement of H-11 and H-12 and  $J = 3.46$  Hz and a *trans* arrangement of H-10 and H-11 with ( $J = 10.30$  Hz). The  $^1H$  NMR chemical shifts of 2 are given in Fig. 1. This is the second report of the neoflavonoid 1 and 2. The next compound isolated from the *C. moonii* leaf extract was identified as apetalactone (3) [1].

The hot petrol extractives of the leaves of *C. walkeri*, when separated on a column of silica gel, gave D:A-friedo-oleanan-3-one (friedelin), 28-hydroxy-D:A-friedo-oleanan-3-one (canophyllol) and an acid mixture. The major component of the latter was found to be calozeylanic acid (4) [7]. We characterized this acid (4) earlier [1] from the leaf extracts of two other *Calophyllum* species, namely, *C. lankaensis* and *C. thwaitesii*. These results are summarized in Table 1.

All the *Calophyllum* species examined had triterpenoids containing the friedo-oleanane skeleton in their leaf extracts. Two major groups of *Calophyllum* species thus emerge. One group (*C. cordato-oblongum* and *C. moonii*) contain neoflavonoids while the other group contain acids in their leaf extracts. Amongst the acid containing group all, except *C. calaba*, had the same acid, calozeylanic acid

\*To whom correspondence should be addressed.

Fig. 1.  $^1\text{H}$ NMR chemical shifts [ $\delta(\text{ppm})$ ] of 1 and 2.

(4). Calozeylanic acid (4) would appear to be the biogenetic precursor of chapelieric acid (7) found in the leaf extract of *C. calaba*.

#### EXPERIMENTAL

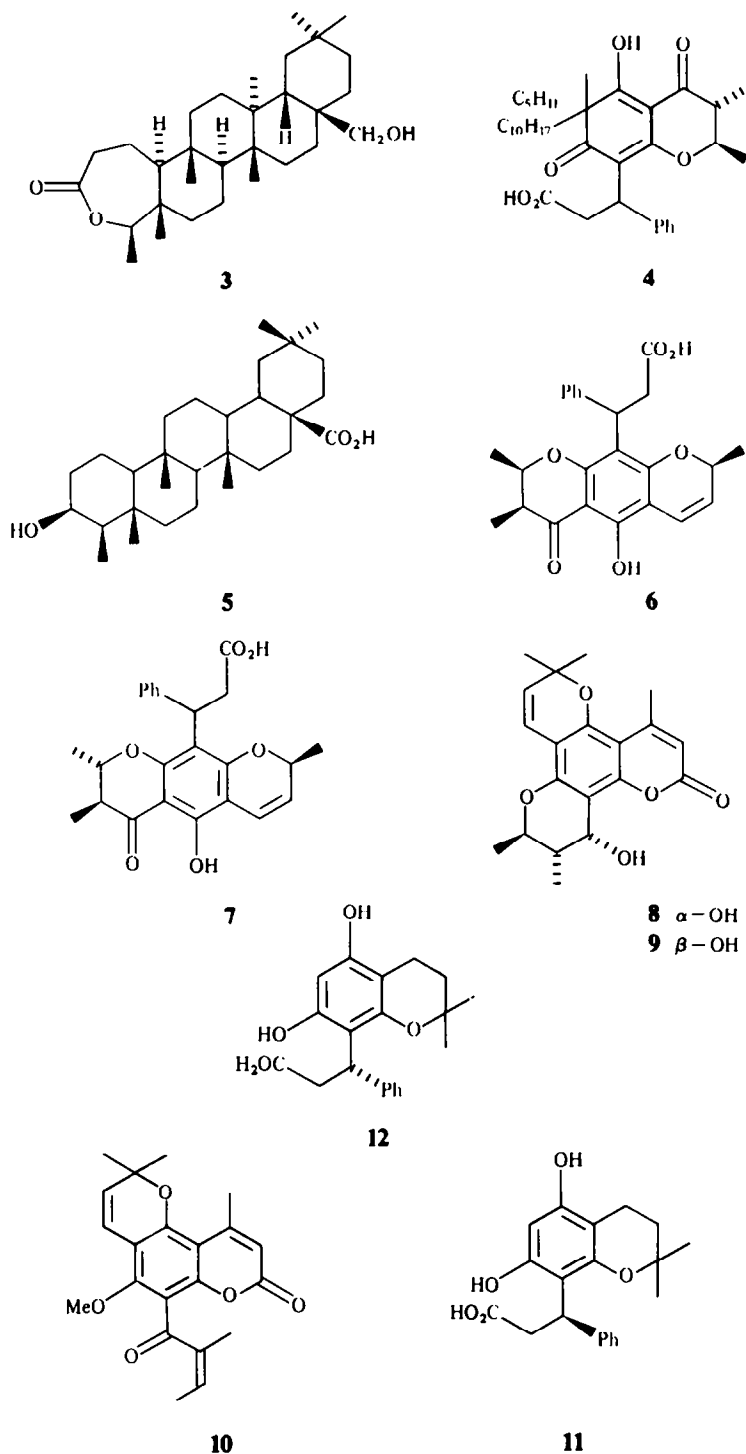
Plants were collected at the Kanneliya rain forest in the South of Sri Lanka.

*Calophyllum moonii* Sun dried milled leaves (4 kg) gave 72 g of hot petrol extract. This was chromatographed over silica gel and elution with petrol-EtOAc gave: sitosterol ester (0.07%) which on hydrolysis (KOH-MeOH) gave sitosterol, mp 137°, lit. [8] 140°;  $[\alpha]_D = -38^\circ$ , lit. [8]  $-35^\circ$ . D:A-Friedooleanan-3-one(friedelin) (0.025%), mp 264–265° lit. [3] 265°;  $[\alpha]_D = -22.5^\circ$  ( $\text{CHCl}_3$ ), lit. [3]  $-22.5^\circ$ . (+)-*cis*-Dihydroinophyllolide (inophyllum A) (1) (0.03%), mp 193–195°, lit. [6] 200°;  $[\alpha]_D =$

$+68.8^\circ$  ( $\text{CHCl}_3$ ), lit. [6]  $+43^\circ$ ; IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3400, 2680–2820, 1710, 1635, 1590, 1445, 1380, 1350, 1150, 850, 770, 700;  $^1\text{H}$ NMR ( $\text{CDCl}_3$ ):  $\delta$  7.30 (5H, m, ArH), 6.51 (1H, d,  $J = 9.90$  Hz), 5.96 (1H, s), 5.33 (1H, d,  $J = 9.90$  Hz), 5.17 (1H, d,  $J = 5.19$  Hz), 4.43 (1H, m,  $J = 3.46$  Hz and 6.92 Hz), 2.27 (1H, m,  $J = 5.19$  and 3.46 Hz), 1.43 (3H, d,  $J = 6.92$  Hz), 1.16 (3H, d,  $J = 7.20$  Hz), 0.98 (3H, s, Me) and 0.97 (3H, s, Me); MS  $m/z$ : 404  $[M]^+$  (25%), 389 (100), 386 (1), 371 (4), 333 (50). Oxidation of 1 with  $\text{CrO}_3$ -pyridine at room temp. gave a product which was purified by prep. TLC to give the oxidized product, mp 155°, lit. [6] 149–151°;  $[\alpha]_D = +65.5^\circ$  ( $\text{CHCl}_3$ ), lit. [6]  $+70^\circ$ ;  $^1\text{H}$ NMR ( $\text{CDCl}_3$ ):  $\delta$  7.40 (5H, m, ArH), 6.58 (1H, d,  $J = 9.90$  Hz), 6.01 (1H, s), 5.42 (1H, d,  $J = 9.90$  Hz), 4.75 (1H, m), 2.60 (1H, m), 1.43 (3H, d,  $J = 7.0$  Hz), 1.16 (3H, d,  $J = 9.0$  Hz), 0.95 (6H, s). (–)-*trans*-Dihydroinophyllolide (soulatrolide) (2) (0.003%), mp 198–200°, lit. [5] 201–202°;  $[\alpha]_D = -21.8^\circ$  ( $\text{CHCl}_3$ ), lit. [5]  $-29.6^\circ$ ; IR  $\nu_{\text{max}}^{\text{CHCl}_3}$   $\text{cm}^{-1}$ : 3100–3500,

Table 1. Distribution of natural products in the leaf extracts of *Calophyllum* species

Species	Triterpenoids	Coumarins	Acids
<i>C. calaba</i>	Friedelin Friedelan-3 $\beta$ -ol Canophyllal Canophyllol Friedelan-3 $\beta$ ,28-diol	—	Canophyllic acid (5) <i>cis</i> -Chapelieric acid (6) Chapelieric acid (7)
<i>C. cordatooblongum</i>	Friedelin Canophyllol	Cordatolide-A (8) Cordatolide-B (9) Oblongulide (10)	—
<i>C. lankaensis</i>	Friedelin Canophyllol Canophyllal Apetalactone (3)	—	Calozeylanic acid (4) Thwaitesic acid (11) <i>iso</i> -Thwaitesic acid (12)
<i>C. moonii</i>	Friedelin Apetalactone (3)	(+)- <i>cis</i> -Dihydroinophyllolide (1) (–)- <i>trans</i> -Dihydroinophyllolide (2)	—
<i>C. thwaitesii</i>	Friedelin Canophyllol Apetalactone (3)	—	Calozeylanic acid (4) Thwaitesic acid (11)
<i>C. trapezifolium</i>	Friedelin Canophyllol	—	Calozeylanic acid (4)
<i>C. walkeri</i>	Friedelin Canophyllol	—	Calozeylanic acid (4)



1370, 1140, 910, 870;  $^1\text{H NMR}$  ( $\text{CDCl}_3$ ):  $\delta$  7.40 (5H, *m*, ArH), 6.55 (1H, *d*,  $J = 10.0$  Hz), 5.96 (1H, *s*), 5.38 (1H, *d*,  $J = 10.0$  Hz), 5.10 (1H, *d*,  $J = 3.46$  Hz), 4.31 (1H, *m*,  $J = 6.29$  and  $10.30$  Hz), 2.8 (1H, OH), 1.80 (1H, *m*,  $J = 3.46$ , 6.77 and  $10.30$  Hz), 1.45 (3H, *d*,  $J = 6.30$  Hz), 1.18 (3H, *d*,  $J = 6.77$ ), 0.93 (6H, *s*,  $2 \times \text{Me}$ ); MS  $m/z$ : 404  $[\text{M}]^+$  (20%), 389 (50), 386 (40), 371 (100), 105 (2), 77 (5). Apetalactone (3) 0.01%, mp 330–332°, lit. [1] 335°;  $[\alpha]_{\text{D}} =$

+33.0° ( $\text{CHCl}_3$ ) lit. [1] +37°, identical with authentic sample (mmp).

*Calophyllum walkeri*. Leaves (2 kg) gave 25 g of hot petrol extract. This on CC over silica gel gave: D:A-friedo-oleanan-3-one (friedelin) (0.01%) mp 262°, lit. [3] 265°;  $[\alpha]_{\text{D}} = -25.5^\circ$  ( $\text{CHCl}_3$ ), lit. [3],  $-22.5^\circ$ . 28-Hydroxy-D:A-friedooleanan-3-one (canophyllol) (0.015%), mp 280°, lit. [3] 280–282°;  $[\alpha]_{\text{D}} = -20^\circ$

(CHCl<sub>3</sub>), lit. [3]  $-21^{\circ}$ . Calozezylic acid (4), (0.5%),  $[\alpha]_D = +14^{\circ}$  (CHCl<sub>3</sub>), lit. [7]  $+12.6^{\circ}$ ; MS *m/z*: 562 [M]<sup>+</sup> (10%), 494 (30), 476 (24), 450 (15), 439 (20), 413 (15), 407 (25), 354 (25), 353 (100), 351 (25) and 345 (12); identical with authentic sample (IR, <sup>1</sup>H NMR, TLC).

**Acknowledgement**—The authors thank the Natural Resources Energy and Science Authority of Sri Lanka for a research grant.

#### REFERENCES

1. Dharmaratne, H. R. W., Sotheeswaran, S. and Balasubramaniam, S. (1984) *Phytochemistry* **23**, 2601.
2. Gunatilaka, A. A. L., De Silva, A. M. Y. J., Sotheeswaran, S., Balasubramaniam, S. and Wazeer, M. I. M. (1984) *Phytochemistry* **23**, 323.
3. Dharmaratne, H. R. W., Sotheeswaran, S., Balasubramaniam, S. and Waight, E. S. (1985) *Phytochemistry* **24**, 1553.
4. Kawazu, K., Ohigashi, H., Takanashi, M. and Mitzui, T. (1972) *Bull. Inst. Chem. Res. Kyoto Univ.* **50**, 160.
5. Gunasekera, S. P., Jayatilaka, G. S., Selliah, S. and Sultanbawa, M. U. S. (1977) *J. Chem. Soc. Perkin Trans. 1*, 1505.
6. Kawazu, K., Ohigashi, H. and Mitsui, T. (1968) *Tetrahedron Letters* 2383.
7. Samaraweera, U., Sotheeswaran, S., Sultanbawa, M. U. S. and Balasubramaniam, S. (1983) *J. Chem. Soc. Perkin Trans. 1*, 703.
8. Samaraweera, U., Sotheeswaran, S. and Sultanbawa, M. U. S. (1983) *Phytochemistry* **22**, 565.